Exotic tree *Artocarpus heterophyllus* (Moraceae) invades the Brazilian Atlantic Rainforest

*Árvore exótica Artocarpus heterophyllus* (Moraceae) invade a Mata Atlântica brasileira

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**Abstract**

*Artocarpus heterophyllus* Lam. (jackfruit) was man made introduced in Tijuca National Park (TNP) in the mid-1800s. Native from Southeast Asia, nowadays densely colonizes the TNP. Here we analyze some jackfruit population parameters that probably allowed the local colonization followed by successful invasion of the new habitat. Based on 20 sample plots, randomly placed inside 5 sites colonized by jackfruit, we described subpopulations hierarchical size diameter structures. Gini’s coefficient values turned around 0.64 and Lorenz’s asymmetry coefficient around 1.03, indicating that size hierarchies were very similar between sites. The five sites were also compared and do not differ based on jackfruit basal area, number of species, Shannon’s index and canopy openness. Jackfruit may, therefore, be considered invasive in all these sites and the low tree diversity values (H’ overall mean 0.74) also indicates that this exotic species locally excludes native ones.

**Keywords:** jackfruit, invasive species, biological invasion, population structure, Tijuca National Park, Brazil.

**Resumo**

*Artocarpus heterophyllus* Lam. (Jaqueira) foi introduzida pelo homem no Parque Nacional da Tijuca (PNT), em meados do século XVIII. Nativa do sudeste da Ásia, esta espécie exótica hoje coloniza densamente o PNT. Neste estudo analisamos alguns parâmetros populacionais da jaqueira que provavelmente permitiram a colonização local e consequentemente a invasão bem sucedida deste novo habitat. A partir da amostragem de 20 parcelas alocadas aleatoriamente em cinco sítios colonizados por jaqueiras, descrevemos as estruturas de diâmetro das subpopulações. Os coeficientes de Gini oscilaram em torno 0,64 e os de Lorenz 1,03, indicando hierarquia de tamanhos semelhante entre as amostras. Os cinco sítios também foram comparados e não diferiram quanto à área basal de jaqueiras, riqueza total de espécies, índice de diversidade de Shannon e abertura do dossel. A jaqueira pode, portanto, ser considerada invasora em todos estes sítios e os baixos valores de diversidade de árvores (H’ em torno de 0,74) também podem indicar que esta espécie exótica exclui localmente as espécies nativas.

**Palavras-chave:** jaqueiras, espécies invasoras, invasão biológica, estrutura de populações, Parque Nacional da Tijuca, Brasil.

**Introduction**

Human-mediated changes in natural ecosystems have caused profound biota modifications, and prominent changes in geographic distributions of plants and animals (Cronk & Fuller 1995; Williamson 1996; Pimentel et al. 2001). Species naturally cross geographical barriers, but human activities such as international commerce and intercontinental travel have strongly contributed to accelerate these transpositions (Richardson et al. 2000; Pimentel et al. 2001). Artificial introductions can be extremely harmful to local ecosystems, causing negative and even unrecognized impacts (Williamson 1996). Mainly because some exotic species are able to change local ecosystem functioning (Kourtev et al. 1999; Zavaleta et al. 2001) and biodiversity patterns (Stohlgren et al. 2001).

Invasive exotic species (IES) also may cause native species extinctions, in some cases comparable to geological mass-extinctions (Cassey et al. 2005),...
leading to biota homogenization (McKinney & Lockwood 1999). World’s biodiversity reduction is straightly related to habitat loss and invasion (Simberloff 2003), both impacts fully experienced by Brazilian ecosystems (e.g., Pivello et al. 1999; Scariot 2001). However, the magnitude of the problems generated by biological invasions will vary depending on the exotic species introduced, on the length of time since its introduction and the susceptibility of the environment to invasion (Williamson 1996).

In Brazil, only a few studies describe IES biology in new habitats, especially trees. Mostly invasion process detection only occurs after high dominance of the invader or another advanced impact on local community can be recognized (Williamson 1996). Since an exotic species is detected colonizing natural habitats, population structure may reveal the capacity of population expanding at present, once this structure can indicate demographic future of species (Bruna & Kress 2002). In this aspect, ancillary tools that describe the degree of inequality in structured populations have been widely used in the description of hierarchies of sizes within populations (Weiner & Solbrig 1984; Solbrig & Solbrig 1984). Among these, we highlight the Gini’s coefficient and the Lorenz’s curve. Gini’s coefficient has been used in biological studies generally to describe size hierarchies and/or intraspecific competition of annual plants (Weiner 1985, 1986; Damgaard & Weiner 2000), system crops (Sadras & Bongiovanni 2004) and tree populations on natural habitats (Rouvinen & Kuuluvainen 2005; Bagchi 2007). On the other hand, the Lorenz curve can be a better descriptor than skewness, generally used to describe population structures (Weiner & Solbrig 1984; Damgaard & Weiner 2000). Furthermore, a population’s stage structure can be the basis for immediate management decisions (Bruna & Kress 2002), specially related to invasive species with long life cycles and increasing populations.

The jackfruit Artocarpus heterophyllus Lam. (Moraceae) is an exotic species native from Southeast Asia, which now occurs widely in reserves of the Atlantic Forest in Rio de Janeiro, Brazilian Southeast. Probably the introduction of jackfruit trees into some areas of the lowest part and the middle slopes of the Tijuca National Park (TNP) began in 1862 with a reforestation program, in which both native and exotic species were simultaneously introduced (Atala et al. 1966; Bandeira 1993; Dean 2002). This program was the first Brazilian initiative to recover a forest ecosystem previously devastated by sugarcane and coffee cycles (Conti et al. 2008). But the widespread information that the whole TNP (4,000 ha) was recovered by that restoration initiative it is not true. Environmental history data of Rio de Janeiro Atlantic forest showed that these reforestation was responsible by 100,000 seedlings planted, covering 170 ha (Oliveira 2007). In fact remaining forest fragments, on higher areas of the Tijuca forest, probably could act as propagule sources colonizing the deforested areas and also enriching the restored sites. Nowadays, in some areas at TNP A. heterophyllus is dominant in density and biomass (Abreu & Rodrigues 2005, Cunha et al. 2006). These populations may have originated from some individuals that were historically introduced in the above-mentioned reforestation, from which the exotic species has reached the status of invasive, since it appears to have expanded naturally.

Furthermore, we compared the population size structures of the invasive tree A. heterophyllus in five sites of TNP. Specifically, we tested the following two predictions: (1) population size structures change among sites, suggesting that some sites are more invasible than others; (2) diversity indexes decrease in places occupied by jackfruits. In testing these hypotheses, our goal is to determine which places of TNP suffered major impacts due to biological invasion, and to set priority sites to management and control of this invasive species.

Material and Methods

Study area

Tijuca National Park (TNP) is one of the largest urban forests in the world, and is widely used as a recreational site (22°55′–23°00′S & 43°11′–43°19′W). Three fragments constitute TNP, subdivided by four sectors with 4,000 ha altogether inside Tijuca Massif, compound by innumerable mountains that reaches until 1021 a.s.l. (IBAMA 2004) situated at the Brazilian second-largest economic and populational center (IBGE 2006). This fact may cause impacts such as uncontrolled urban growth in the boundary regions (Coelho-Netto et al. 2007), atmospheric pollution (Azevedo et al. 1999) and the presence of non-native fauna (Cunha et al. 2006) and flora (Bandeira 1993).
Jackfruit invades the Atlantic Rainforest

The annual precipitation varies from 1300 mm to slightly more than 3000 mm, with abundant precipitation also in winter (Mattos et al. 1970) and torrential rains in the hot summer months (December-February) (Coelho-Netto et al. 2007). Lithosols, cambisols and latosols predominate in the upper part, and podzolic soils in the lower part (Coelho-Netto et al. 2007).

The TNP vegetation is typical of tropical rain forests, characterized by big trees, palms, ferns, epiphytes, and lianas. In the mature forest, emergent trees reach 25 m height and canopy strata mean height is about 20 m (Atala et al. 1966). Within the TNP, there is a mosaic of vegetation ranging from undisturbed forests to patches totally invaded by African grasses. Notwithstanding, the most representative families in that study were Myrtaceae, Lauraceae, Leguminosae, Rubiaceae, Euphorbiaceae, Meliaceae and Annonaceae. The more representative species were Coussarea noddosa (Benth.) Müll. Arg., Geonoma schottiana Mart., Myrcia rostrata DC., Myrcia laxiflora Cambess., Allophylus edulis (A. St.-Hil., Cambess. & A. Juss.) Radlk., Cryptocarya moschatana Nees & C. Mart., Tetrorchidium aff. rubrivenium Poepp., Eugenia prasina O. Berg, Pausandra megalophylla Müll. Arg. and Trigynaea sp. Schltdl. (Oliveira et al. 1995).

The studied species

Artocarpus heterophyllus Lam. is a species originated from Southeast Asia (Chaves et al. 1967). In the year 1682, in the state of Bahia, there were already 11 jackfruit trees of unknown origin. The first record of a shipment to Brazil dates from January 1683 (Ferrão 1993). The recently arrived seeds and seedlings were sent to a nursery in Bahia State to be acclimated, and were later distributed to other regions of the country (Dean 2002). In 1803 in Rio de Janeiro municipality, naturalized adult individuals apparently already existed (Chaves et al. 1967).

The jackfruit is a large tree, reaching a height of more than 10 meters; the trunk may exceed 1 meter in diameter. It is monoeccious and cauliflorous (Chaves et al. 1967; Crane et al. 2002). The species can be pollinated by insects or wind, tolerates climate stresses well, and in ideal orchard conditions it can fruit in three or four years (Crane et al. 2002). In Brazil, a well-developed jackfruit tree can produce up to 100 fruits per year, some weighing more than 30 kg (Chaves et al. 1967). Individuals of this species fruit asynchronously during the entire year, with a large proportion preferentially fruiting in summer (December-February). The fruits are consumed by humans and by other mammals such as rodents and primates (Cunha et al. 2006). Field observations indicated a high germination rate of seeds in edge areas or where the species already occurred (R.R. Oliveira, personal communication). Presently, its distribution includes nearly the entire territory of Brazil in a fairly wide range of habitats, colonizing open and forested areas associated with human-impacted environments (Caraúra & Díaz 2002).

In Rio de Janeiro, it densely colonizes edge areas of the Atlantic Forest, and is found in Reserves such as the Tijuca National Park and the Poço das Antas, União and Tinguá biological reserves (Abreu & Rodrigues 2005).

On the native habitat (Southeast Asia), A. heterophyllus is typical of advanced successional stages, and grows naturally up to an altitude of 1300 m in the sub-canopy of the Monodominant Humid Tropical Forest. The species is considered rare (up to 1 individual per ha) in this environment, where few species dominate the different forest strata (Chittibabu & Parthasarathy 2000; Bhuyan et al. 2003). It is shade-tolerant, and germinates best in clearings; the fruits mature between July and August, and the seeds vary in size and weight (1.5 to 14 g). Their dispersers include rodents, monkeys and wild pigs (Khan 2004). The seeds have a relatively high rate of predation (Sodhi et al. 2003) and germinate in the rainy season, during the summer (May-September) when 85% of the annual precipitation occurs (Chittibabu & Parthasarathy 2000; Bhuyan et al. 2003). In this manner, the seeds germinate and the seedlings establish themselves, becoming able to easily tolerate the December-February dry season (Chittibabu & Parthasarathy 2000; Bhuyan et al. 2003).

Sites descriptions

Five study sites were sampled at TNP. They have similar disturbance histories, because comprised areas which were at one time deforested for sugarcane and/or coffee cultivation, and where the jackfruit trees were later introduced and naturally disseminated. Random sampling methods were used to demarcate plot locations at each site (Mueller-Dombois & Ellenberg 1974).

Site 1 – Located in the “A” sector of TNP, a fragment with approximately 1,300 ha. This site is locally known as “Represa dos Ciganos”. Six plots were
placed at Northern exposed slopes. This fragment has a well structured forest with tall trees and there were the tallest jackfruit trees.

Site 2 – Locally known as “Horto Florestal”, that site belongs to the “B” sector of TNP, the bigger fragment with approximately 1,660 ha. Two plots were placed close to the well preserved forest remnant known as “Mata do Pai Ricardo”, located at Southern slope near a boundary region. At that place, jackfruit densely colonized roadsides and it is believed that jackfruit root systems can prevent sliding of slopes.

Site 3 – This site is located in the “C” sector, which is a forest fragment with high hills, covering about 230 ha. Site 3 comprises four plots placed on the Southern slopes. Two from these plots were placed in the riparian area.

Site 4 – It is also located in the “C” sector, but plots were established on the Northern slopes.

Site 5 – This site is locally known as “Parque Lage”, which was a sugar cane and coffee farm in the past, at present covered by secondary forest and planted trees. Since 2004 it was embedded to the “B” sector of TNP. The vegetation structure is very similar to the other areas of TNP but exotic species occur at the bottom regions. Four plots were located close to a trackside used by park visitors (track to “Corcovado”), exposed to Southern direction.

Structures of trunk diameters

In the year 2004, 20 circular permanent plots (10 m radius; 314 m² each) were set to survey jackfruit populations at TNP. The 20 plots were distributed in five sites scattered throughout TNP, where jackfruit populations were recorded.

All individuals of the plant community with a diameter at breast height (DBH) ≥ 5 cm were then marked and measured. In the year 2009, all those individuals were again measured and individuals with DBH ≥ 1 cm were included in the survey. To calculate individual basal areas (BA) we used the formula: \( S = \pi r^2 \); where: \( S \) = individual basal area (circle area); \( \pi = 3.14 \) and \( r = \) radius (radius = half of the DBH measured). Individuals with visibly interlinked multiple trunks were considered as a single individual; in these cases, each trunk was measured separately and their BA were summed, and from the resulting basal area a single virtual diameter > 1 cm was obtained.

The basal areas of all individuals were calculated from the DBHs. One-Way ANOVAs followed by an *a posteriori* Tukey’s test (HSD for unequal Ns; \( p < 0.05 \)) were used to assess whether the parameters analyzed (number of individuals, total basal area, basal area of jackfruits) differed significantly among sites, because any difference found may be indicative of different processes and patterns experienced by each subpopulation. The site 2 was not considered due to small number of replicates (2).

To test the hypothesis that (1) population size structure change among sites, suggesting that some sites are more severely invaded than others, we performed trunk diameters structure from the five sites and made comparisons between than through frequency distributions, measures for size inequalities in population structure, and spatial distribution at plot scales.

Attempts to confirm the age of these individuals by their wood anatomy were unsuccessful, since this species does not form growth rings.

**Frequency distribution**

Diameter classes were established according to morphological characters observed in the field during the pilot study to define ontogenetic stages (e.g., presence of reproductive structures and characteristics of the crown and/or trunk). A frequency histogram were developed, where the class JUVENILES 1 was composed of small sized trees with DBH ranging from 1 to 4.9 cm; JUVENILES 2 was composed of small- to medium-sized trees with DBH ranging from 5 to 9.9 cm and the crown little developed. The individuals of class JUVENILES 3 included small to medium trees with DBH ranging from 10 to 14.9 cm, still with the crown relatively little developed. The class of PRE-REPRODUCTIVES included individuals from 15 to 24.9 cm DBH, composed of large immature (non-reproductive) trees, with a relatively full crown. The reproductive individuals were divided into the classes ADULTS 1 and 2. The class ADULTS 1 included trees with DBH from 25 to 44.9 cm, with a smooth trunk. The class ADULTS 2 included individuals with a diameter equal or greater than 45 cm, generally with bark fissures on the trunk.

**Population inequality measures**

Size inequality was assessed using Lorenz’s curves, Gini’s coefficient (Weiner & Solbrig 1984), Lorenz’s asymmetry coefficient (Damgaard & Weiner 2000) and their respective confidence intervals. In the Lorenz’s curve, individuals were ranked according to their size (DBH in this case)
Spatial distribution

The spatial distribution pattern was determined using Morisita’s index. To calculate this index all 20 sample plots were considered. As observed by Souza & Martins (2002), the Morisita Index ($I_m$) gives the type of spatial distribution, indicated by the values: $I_m = 1$ random distribution, $I_m > 1$ aggregated distribution, and $I_m < 1$ uniform distribution, and a measurement of the degree of aggregation, whereas other measurements of dispersion test only one of these patterns. This index has desirable statistical properties: it has a known sampling distribution, and is not influenced by sample size. To test the significance of the values of $I_m$ of 1.0, the chi-square test was used: $\chi^2 (df = Q-1) = (Q-1) s^2 / \mu$ (Souza & Martins 2002 apud David & Moore 1954), where $Q$ is the total number of plots, and $s^2$ and $\mu$ are, respectively, the variance and mean of the number of jackfruit trees per plot.

Richness, biodiversity and canopy structure

Community sampled trees were classified in morphospecies. From these data, PC-ORD 5.19 (McCune & Mefford 1999) was used to calculate richness and Shannon-Weiner biodiversity index in every plot.

To describe canopy structure, we calculate canopy openness after taking digital pictures with a Nikon Coolpix 990, a fisheye lens (Nikon FC-08) and a tripod one meter high. Pictures were taken according to terrain slope in the center of plots during cloudy homogeneous skies from 2007 summer. To analyze pictures we used WINSCANOPY 2003.d in an automatic threshold manner (Regent 2003).

Comparisons among sites were done after normality tests. Since variables do not fit to normal distribution, Kruskal-Wallis tests were applied. Site 2 was not included in the analyses due to small number of replicates (2).

Comparisons with native species

In order to detect population patterns of native species to compare to the populations of jackfruits, secondary data from phytossociological samples in areas of the Atlantic Forest were used, in which only those species with the highest values of basal area were selected.

Results

Within the plots, the subpopulations contained high relative densities of jackfruits, ranging from 40% to 100% of the individuals in a locality. A total of 1262 JUVENILES 1 (DBH 1 to 4.9 cm) and 564 bigger trees (DBH > 5 cm) were sampled in 2009, and the mean basal area of jackfruits was 1.712 ± 0.773 m² plot⁻¹, with approximately 28 ± 11 trees plot⁻¹ (Tab. 1).
The structure of the jackfruit population in TNP in 2009 (diameters histogram) showed the right-asymmetrical log-linear distribution pattern, known as the “reverse J” (y=182.4e-0.5138x; R²=0.72). The largest proportion of the individuals (70%) were in the first size class, and the classes of reproductive adults corresponded to 9% of the individuals of the population.

The densest subpopulations contained many immature and few adults. However, as expected, adults were responsible for the greater proportion of the basal area, which were distributed unequally between adults and immatures (Tab. 2).

Maximum DBH ranged from 80.9 in site 1 to 108.7 in site 3. Gini’s coefficients were all near 0.65 and Lorenz’s asymmetry coefficients were all near 1 (Tab. 3). These values indicate high similarity in size hierarchies and no sample asymmetry were observed between smaller and bigger individuals in a single site. No differences were observed between sites. Analyzing the confidence intervals of the coefficients, all sites had the same size hierarchies. In the landscape scale, Artocarpus heterophyllus showed clumped patterns, resulting in a Morisita’s Dispersion Index value of 1.11.

The sites were also very similar in terms of basal area (BA), richness (S), Shannon-Wiener diversity index (H’) and canopy openness (Tab. 4). Mean basal areas were about 1.71 (± 0.77) m² plot⁻¹, mean richness was 6 (± 4) species plot⁻¹, H’ was about 0.74 and mean canopy openness was 6.16 %.

Comparing the basal area (BA) of the sub-populations of A. heterophyllus with the BA of other species of the Atlantic Forest (Tab. 5), the BA occupied by jackfruits was 10.9 to 98.1 times larger than the BA of native tree species of the Atlantic Forest.

Discussion
Although exotic species may be rare in undisturbed tropical forests, in disturbed forests such species may become invasive and even cause drastic changes to the ecosystem, including indirect habitat destruction, facilitating their own dominance over the native species (Fine 2002). Jackfruit nowadays densely colonizes many sites at Tijuca National Park (TNP), an urban and permanently disturbed forest remnant. In addition, impacts on natural habitats, such as fragmentation and habitat loss, may favor the dissemination of this exotic species (Davis et al. 2000; Fine 2002). In this way, the successful man-mediated introduction of jackfruits into the TNP (Atala et al. 1966; Bandeira 1993) after 150 years resulted in an effective habitat invasion that probably represents the most massive invasion promoted by a tree species at Atlantic Rainforest.

Other Brazilian Reserves that have edge sites colonized by jackfruit showed an increased basal area when compared to other edge sites (Rodrigues 2004). This fact may indicate that even populations of native species with larger basal areas in local communities are incapable to colonize the locality in the same manner as the jackfruits. The jackfruit colonization success as well as the dominance in density and biomass (Abreu & Rodrigues 2005; Cunha et al. 2006) can be also favored by population patterns such as clumped distribution, dispersal way (barochory), and by the probably escape from natural enemies (Sax & Brown 2000; Keane & Crawley 2002). The time elapsed since the introduction of the species by planting in a locality may also contribute for species dissemination (Williamson 1996). Furthermore, the jackfruit seed predation rate is high in preserved areas and low in forest edges, and in the latter areas the species shows a high germination (R.R. Oliveira, personal communication). In a pilot study (unpublished data) we observed single plots (2 m²) having more than 200 jackfruit seedlings. This shows that the jackfruit can regenerate better than other species from the Atlantic Rainforest. And it can be explained because in tropical forests, plants with larger seeds tend to germinate more readily than plants with small seeds (Dawson et al. 2009).

Table 1 – Summary of parameters used to characterize the plant community structure in the plots studied in Tijuca National Park in 2009 (DBH ≥ 1 cm). BAl = entire community (m² ha⁻¹) basal area; BAm = basal area composed exclusively of jackfruit in the community (m² ha⁻¹); BAm = basal area occupied by other individuals, not jackfruits (m² ha⁻¹); N = number of individuals in the community (trees ha⁻¹); N = number of jackfruit individuals in the community (trees ha⁻¹). Mean ± SD = mean and standard deviation of parameters by plot (314 m²).

<table>
<thead>
<tr>
<th></th>
<th>BAl (m² ha⁻¹)</th>
<th>BAm (m² ha⁻¹)</th>
<th>BAm (m² ha⁻¹)</th>
<th>N (trees ha⁻¹)</th>
<th>N (trees ha⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total</td>
<td>68.446</td>
<td>54.351</td>
<td>15.916</td>
<td>1107</td>
<td>889</td>
</tr>
<tr>
<td>Mean ± SD</td>
<td>2.149 ± 0.787</td>
<td>1.649 ± 0.762</td>
<td>0.952 ± 2.115</td>
<td>34.75 ± 9.5</td>
<td>27.9 ± 11.5</td>
</tr>
</tbody>
</table>
Table 2 – Jackfruit demographic patterns of the study sites (mean ± standard deviation). Sites = localities (number of sites in the locality, see methods text); Immatures = individuals with DBH ranging from 5 to 24.9 cm; Adults = individuals with DBH greater than 25 cm; n = number of individuals; % n = percentage of number of individuals; BA = basal area; % BA = percentage of basal area; ANOVA = values of MS for One-Way ANOVAs, df = 15, p < 0.05, Tukey’s test for unequal Ns. Symbol (*) indicates that no significant differences were observed.

<table>
<thead>
<tr>
<th>Sites n</th>
<th>Immatures</th>
<th></th>
<th>Adults</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>% n</td>
<td>BA</td>
<td>% BA</td>
<td>n</td>
</tr>
<tr>
<td>1 (6)</td>
<td>23 ± 10</td>
<td>71</td>
<td>0.28 ± 0.14</td>
<td>17</td>
</tr>
<tr>
<td>2 (2)</td>
<td>25 ± 13</td>
<td>79</td>
<td>0.34 ± 0.19</td>
<td>19</td>
</tr>
<tr>
<td>3 (4)</td>
<td>16 ± 7</td>
<td>73</td>
<td>0.16 ± 0.07</td>
<td>10</td>
</tr>
<tr>
<td>4 (4)</td>
<td>15 ± 5</td>
<td>61</td>
<td>0.19 ± 0.11</td>
<td>13</td>
</tr>
<tr>
<td>5 (4)</td>
<td>22 ± 11</td>
<td>73</td>
<td>0.22 ± 0.10</td>
<td>12</td>
</tr>
<tr>
<td>ANOVA*</td>
<td>82.889</td>
<td>0.01498</td>
<td>17.167</td>
<td>0.63681</td>
</tr>
</tbody>
</table>

Table 3 – Parameters analyzed to describe the structure hierarchy of sizes (DBH) in the sites studied in the Tijuca National Park, RJ (2009). Site = studied site; n = number of individuals; mean ± SD = mean and standard deviation of DBH; max DBH = greater DBH observed in the site; G’ = corrected Gini’s coefficient; CI G’ 95% = confidence interval for G’; S = Lorenz’ Asymmetric Coefficient; CI S 95% = confidence interval for S.

<table>
<thead>
<tr>
<th>Site</th>
<th>n</th>
<th>DBH max</th>
<th>DBH</th>
<th>G</th>
<th>CI G’ 95%</th>
<th>S</th>
<th>CI S 95%</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>723</td>
<td>6.77 ± 11.42</td>
<td>80.9</td>
<td>0.6452</td>
<td>[0.6232 - 0.6615]</td>
<td>1.0346</td>
<td>[1.0006 - 1.0794]</td>
</tr>
<tr>
<td>2</td>
<td>196</td>
<td>7.80 ± 13.07</td>
<td>81.9</td>
<td>0.6481</td>
<td>[0.5931 - 0.6815]</td>
<td>1.0375</td>
<td>[0.9497 - 1.0977]</td>
</tr>
<tr>
<td>3</td>
<td>205</td>
<td>10.41 ± 17.39</td>
<td>108.7</td>
<td>0.6467</td>
<td>[0.5891 - 0.6780]</td>
<td>1.094</td>
<td>[0.9890 - 1.1470]</td>
</tr>
<tr>
<td>4</td>
<td>298</td>
<td>9.15 ± 10.03</td>
<td>83.6</td>
<td>0.6551</td>
<td>[0.6328 - 0.6736]</td>
<td>0.9682</td>
<td>[0.9050 - 1.0304]</td>
</tr>
<tr>
<td>5</td>
<td>404</td>
<td>7.46 ± 13.51</td>
<td>107.7</td>
<td>0.6459</td>
<td>[0.6061 - 0.6753]</td>
<td>1.0317</td>
<td>[0.9843 - 1.0954]</td>
</tr>
</tbody>
</table>

Table 4 – Means (± SD) from parameters used in the comparisons between the sites. DBH 1 = Basal area composed of jackfruits with DBH ≥ 1 cm; DBH 10 = Basal area composed of jackfruits with DBH ≥ 10 cm; S = species richness; H = Shannon-Weiner biodiversity index; Openness = canopy openness (%). No significant differences were observed between sites.

<table>
<thead>
<tr>
<th>Site</th>
<th>DBH 1</th>
<th>DBH 10</th>
<th>S</th>
<th>H</th>
<th>Openness</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1.67 ± 0.99</td>
<td>1.58 ± 0.97</td>
<td>4.33 ± 2.07</td>
<td>0.53 ± 0.45</td>
<td>5.64 ± 0.48</td>
</tr>
<tr>
<td>2</td>
<td>1.78 ± 0.00</td>
<td>1.70 ± 0.02</td>
<td>6.00 ± 1.41</td>
<td>0.72 ± 0.44</td>
<td>5.88 ± 0.35</td>
</tr>
<tr>
<td>3</td>
<td>1.65 ± 0.90</td>
<td>1.59 ± 0.86</td>
<td>4.25 ± 3.59</td>
<td>0.75 ± 0.80</td>
<td>6.16 ± 0.34</td>
</tr>
<tr>
<td>4</td>
<td>1.64 ± 0.88</td>
<td>1.59 ± 0.87</td>
<td>9.75 ± 6.24</td>
<td>1.18 ± 0.92</td>
<td>7.66 ± 1.33</td>
</tr>
<tr>
<td>5</td>
<td>1.89 ± 0.72</td>
<td>1.80 ± 0.70</td>
<td>5.25 ± 1.71</td>
<td>0.62 ± 0.05</td>
<td>5.60 ± 0.70</td>
</tr>
</tbody>
</table>

Ecosystems invasibility must not be generalized, because they are closely dependent on scale, vegetation type, biome, and resources availability (Stohlgren et al. 1999). Furthermore, in some cases it is not necessary to the invader to be aggressive when this species arrived to occupy a potential available niche (Heger & Trepl 2003). Jackfruit that colonizes high biodiversity Atlantic Forest areas like TNP, at first excludes local plant species. In this case, the invasion success was determined by both the attributes of the invasive species and the susceptibility of the invaded ecosystem (Williamson 1996; Heger & Trepl 2003). Additionally, disturbance and propagule pressure are factors that deserve attention because although they are confused, both factors play important roles in invasion of tropical forests (Edward & Munishi 2009).

Jackfruit in his original habitat is rare, experiences a monsoon seasonally climate condition (Ayyappan & Parthasarathy 1999;
Table 5 – Secondary data extracted from phytosociological surveys in Atlantic Forest areas in Rio de Janeiro. DBH = minimum diameter adopted as inclusion criteria; P (ha) = size of original sampling area in hectares; BA = basal area (m²) occupied by the species extrapolated to hectares; source = reference from phytosociological survey.

<table>
<thead>
<tr>
<th>Species</th>
<th>DBH (cm)</th>
<th>P (ha)</th>
<th>BA (m² ha⁻¹)</th>
<th>Source and place</th>
</tr>
</thead>
<tbody>
<tr>
<td>Artocarpus heterophyllus Lam.</td>
<td>5</td>
<td>0.63</td>
<td>53.68</td>
<td>This study*</td>
</tr>
<tr>
<td>Artocarpus heterophyllus</td>
<td>10</td>
<td>0.63</td>
<td>52.25</td>
<td>This study*</td>
</tr>
<tr>
<td>Metrodorea brevifolia Engl.</td>
<td>10</td>
<td>1</td>
<td>4.13</td>
<td>Silva &amp; Nascimento (2001)</td>
</tr>
<tr>
<td>Paratecoma peroba (Record &amp; Mell) Kuhlm.</td>
<td>10</td>
<td>1</td>
<td>1.26</td>
<td>Tabuleiros do Norte - RJ</td>
</tr>
<tr>
<td>Pseudopiptadenia contorta (DC.)</td>
<td>10</td>
<td>1</td>
<td>0.57</td>
<td>GP Lewis &amp; M.P. Lima</td>
</tr>
<tr>
<td>Trichilia pseudostipularis (A. Juss.) C. DC.</td>
<td>10</td>
<td>1</td>
<td>0.54</td>
<td></td>
</tr>
<tr>
<td>Hyeronima alchorneoides Allemão</td>
<td>10</td>
<td>0.6</td>
<td>4.26</td>
<td>Moreno et al. 2003*</td>
</tr>
<tr>
<td>Pseudopiptadenia contorta</td>
<td>10</td>
<td>0.6</td>
<td>2.15</td>
<td>Região do Imbé - RJ</td>
</tr>
<tr>
<td>Rustia formosa (Cham. &amp; Schltdl. ex DC.) Klotzsch</td>
<td>10</td>
<td>0.6</td>
<td>2.13</td>
<td></td>
</tr>
<tr>
<td>Virola oleifera (Schott) A.C. Sm.</td>
<td>10</td>
<td>0.6</td>
<td>2.10</td>
<td></td>
</tr>
<tr>
<td>Actinostemon verticillatus (Klotzsch) Baill.</td>
<td>10</td>
<td>0.6</td>
<td>1.60</td>
<td>Moreno et al. 2003*</td>
</tr>
<tr>
<td>Mabea fistulifera Mart.</td>
<td>10</td>
<td>0.6</td>
<td>1.57</td>
<td>Região do Imbé - RJ</td>
</tr>
<tr>
<td>Maytenus commutata Reissek</td>
<td>10</td>
<td>0.6</td>
<td>1.53</td>
<td></td>
</tr>
<tr>
<td>Vochysia oppugnata (Velloso) Warm.</td>
<td>10</td>
<td>0.6</td>
<td>1.47</td>
<td></td>
</tr>
<tr>
<td>Cupania emarginata Cambess.</td>
<td>10</td>
<td>0.4</td>
<td>4.81</td>
<td>Rodrigues 2004*</td>
</tr>
<tr>
<td>Ecclinusa ramiflora Mart.</td>
<td>10</td>
<td>0.4</td>
<td>3.08</td>
<td>REBIO União - RJ</td>
</tr>
<tr>
<td>Ficus gomelleira Kunth &amp; C.D. Bouché</td>
<td>10</td>
<td>0.4</td>
<td>2.93</td>
<td></td>
</tr>
<tr>
<td>Pradosia kuhlmannii Toledo</td>
<td>10</td>
<td>0.4</td>
<td>2.87</td>
<td></td>
</tr>
<tr>
<td>Virola bicuhyba (Schott ex Spreng.) Warb.</td>
<td>10</td>
<td>0.4</td>
<td>2.78</td>
<td></td>
</tr>
<tr>
<td>Alchornea triplinervia (Spreng.) Müll. Arg.</td>
<td>5</td>
<td>1</td>
<td>5.1</td>
<td>Guedes-Bruni et al. 1997</td>
</tr>
<tr>
<td>Cabrlea canjerana (Vell.) Mart.</td>
<td>5</td>
<td>1</td>
<td>1.2</td>
<td>Macaé de Cima - RJ</td>
</tr>
<tr>
<td>Euterpe edulis Mart.</td>
<td>5</td>
<td>1</td>
<td>1.9</td>
<td></td>
</tr>
<tr>
<td>Mollinedia gigliana Perkins</td>
<td>5</td>
<td>1</td>
<td>1.5</td>
<td></td>
</tr>
<tr>
<td>Myrcia pubipetala Miq.</td>
<td>5</td>
<td>1</td>
<td>1.4</td>
<td></td>
</tr>
<tr>
<td>Casearia obligna Spreng.</td>
<td>5</td>
<td>1</td>
<td>1.6</td>
<td>Pessoa et al. 1997</td>
</tr>
<tr>
<td>Croton floribundus Spreng.</td>
<td>5</td>
<td>1</td>
<td>2.6</td>
<td>Macaé de Cima - RJ</td>
</tr>
<tr>
<td>Euterpe edulis</td>
<td>5</td>
<td>1</td>
<td>1.9</td>
<td></td>
</tr>
<tr>
<td>Nephelea setosa (Kaulf.) R.M. Tryon</td>
<td>5</td>
<td>1</td>
<td>1.2</td>
<td></td>
</tr>
<tr>
<td>Tibouchina scrobiculata Cogn.</td>
<td>5</td>
<td>1</td>
<td>3.9</td>
<td></td>
</tr>
</tbody>
</table>

* extrapolated values for hectares for comparison purposes only. The species were not observed in a continuous hectare.

Parthasarathy (1999) and exhibit synchronicity in fruiting processes (Khan 2004). Otherwise, in the Brazilian Atlantic Forest the absence of a marked dry season (Mattos et al. 1970) with water supply year-round a continued reproduction may be favored. In fact, it is expected that exotics grow faster and reproduce better in some invaded environments (Jakobs et al. 2004). It was also observed in TNP that jackfruits do not exhibit synchronicity in relation to the fruiting period, even considering the reproducing peak at summer. That is, different individuals, even neighbors, fruit at different times of the year. This is a common pattern in species.
of the family Moraceae (J.P.P. Carauta, personal communication), as often observed in fig species (Romo 1996; Korine et al. 2000; Wendeln et al. 2000).

Asymmetric competition increases variation in growth rates between dominant and suppressed plants, its onset promotes size inequality, and this effect is heightened by time (Crawley 1997). Even with an uncertain elapsed time after species first establishment at each site, 150 years after introduction in TNP, population characteristics give to the jackfruit the same size hierarchy and a very similar population structure at different sites, showing that the biological invasion process is very similar in all sites. Jackfruit showed a population structure typical from shade-tolerant species, with many individuals in the first size class (JUVENILE 1). Together, the biology of a species and structures of diameters can provide valuable information about the successional process undergone by a subpopulation (White 1980). Different size classes may compose a stable population, and the proportions of these classes will vary in accordance to the abiotic and biotic factors those population individuals and their ancestors experienced historically (Sarukhán 1980; Hutchings et al. 2000). Trees are more likely to survive if their neighbors die (Crawley 1997), so invasive species can supplant native species as the native die.

The barochory dispersal syndrome favors clumped spatial patterns as observed in these populations of jackfruit. That spatial pattern, especially of young and pre-reproductive individuals, has also been observed for native tree species of the Atlantic Forest such as *Miconia cinnamomifolia* (DC.) Naudim (Pereira 1998), *Calophyllum brasiliense* Camb. (Marques & Joly 2000) and *Caesalpinia echinata* Lam. (Rodrigues et al. 2009). Pereira (1998) characterized *M. cinnamomifolia* as an initial secondary species, because it grows most densely in edge sites, although its density decreases as ecological succession progresses. In their native environment, jackfruits are considered typical of advanced successional stages (Khan 2004). In the Atlantic Forest, however, this species has occupied areas of secondary vegetation, from open areas to dense forests.

In the case of the jackfruits at TNP, both the local man-mediated introduction and the individual longevity explain the presence of this exotic per se. Although from some historically introduced individuals the species was capable of expanding its subpopulations, becoming invasive in areas of the Atlantic Forest underwent some disturbance (e.g., park boundaries, roadsides). Considering that no differences were found among sites, based on basal area, subpopulation hierarchies, richness, biodiversity and canopy openness, it is probable that jackfruit is potentially an invasive species at TNP, even in places with small populations at present. All data showed that the Brazilian Atlantic Rainforest is a new habitat that allows the jackfruit to proliferate and dominate the plant community. So any place where jackfruit colonizes native forests in this ecological region can be a priority area for management, eradication being recommended at the first detection.

Acknowledgments

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